

Development and validation of LC/MS/MS method for determination of mycotoxins

Tijana Serafimovska¹, Marija Darkovska Serafimovska², Marija Mitevska³, Gjoshe Stefkov¹, Jasmina Tonic Ribarska¹

¹Faculty of Pharmacy, University Ss.Cyril and Methodius, address: Mother Tereza 47, 1000 Skopje, North Macedonia, <u>ff.ukim.edu.mk</u>
 ² Faculty of Medical sciences, University Goce Delcev, address:Krste Misirkov No.10-A P.O. Box 201, 2000 Shtip, North Macedonia, ugd.edu.mk
 ³ NYSK Holdings, address: Cojlija, Street 112, No.127, 1000 Skopje, Republic of North Macedonia, nyskholdings.com.mk

INTRODUCTION

Mycotoxins (aflatoxins and ochratoxin A) are secondary toxic metabolites, that contaminate raw materials that are usually used in the preparation of products for human use. Presence of these contaminants in the preparation for human use, can causes various acute and chronic impacts on human health. Carcinogenicity,

MATERIAL AND METHODS:

Chemicals and Regents

Liquid standards of aflatoxin B1, aflatoxin B2, aflatoxin G1, aflatoxin G2 and ochratoxin A, as well as Immuno-affinity columns were supplied by R-biopharm (Germany).

Apparatus: Liquid chromatography was performed on LC/MS/MS system from Shimadzu.

Chromatographic Conditions and analyte transitions are given in Table 1 and Table 2

Column Raptor Biphenyl 100 mm x 2.1 mm, particle size 2.7 µm, (Cat.No.980-18088)

Guard Raptor Biphenyl EXP Guard Column Column Cartridge 2.7 µm 5 x 2.1 mm

RESULTS:

Validation of the method

The calibration characteristics and validation parameters are shown in Table 3. Linearity of response was calculated as a ratio of peak areas of Aflatoxins B1, B2, G1, G2 and Ochratoxin A in standard solution vs. concentration in spiked samples in the concentration range of $0.1 - 5\mu g/L$ for Aflatoxin B1, B2, G1 and G2 and for Ochratoxin A from $1 - 50\mu g/L$. Coefficient of correlation was greater than 0.999 for all mycotoxins.

Results from the limit of detection/limit of quantification for mycotoxins and precision and accuracy of the method are shown in Table 4 and Table 5.

	AfG2	AfG1	AfB2	AfB1	OchA		Concentration added	Measured (µg/L) ^a	concentration
Linearity		01	5 (µg/L)	1–50 (µg/L)		AfG2	(~5/-)	Recovery
range		0.1	σ (μg/ L)			1.5 (µg/L)	$1.215 \pm 0.96\%$	81.0%
Determ.							$2.0 (\mu g/L)$	1.892 ± 0.82%	94.6%
	0.999	0.999	0.999	0.999	0.999		5.0 (µg/L)	4.321 ± 0.78%	86.42%
						F	AfB2		Recovery
							$1.5 (\mu g/L)$	$1.228 \pm 0.87\%$	81.86%
Table 3. Characteristics of the linear regression analysis					ression analysis		2.0 (µg/L)	$1.927 \pm 0.58\%$	96.35%
							$5.0 (\mu g/L)$	4.283 ± 0.72%	85.66%
							AfG1		Recovery
Mycotoxi	n Lin	nit of d	letection	n	Limit of		1.5 (µg/L)	1.214± 0.58%	80.9%
					uantification		2.0 (µg/L)	1.940± 0.82%	97.0%
							5.0 (µg/L)	4.696± 0.85%	93.9%
AflG2		0,023			0,069	E .	AfB1		Recovery
AflG1		0,017			0,053		$1.5 (\mu g/L)$	1.334± 0.71%	88.93%
		·					$2.0 (\mu g/L)$	$1.938 \pm 0.49\%$	96.9%
AflB2	0,034			0,105	free a	5.0 (µg/L)	4.900± 0.57%	98.0%	
AflB1	0,027			0,082		OchA		Recovery	
OchA		0,32	29		0,997		$15 (\mu g/L)$	14.09± 0.86%	93.93%
Comr					0,777		$\frac{20 (\mu g/L)}{1000000000000000000000000000000000000$	20.93±0.93%	104.6%
Table 4. L	OD/L	OQ of	mycoto	xins			50 (μg/L)	50.59± 1.03%	101.18%
							Table 5. Precisio	n and accuracy	of the method
225000-1								Max Time 12.372 Scan# 3,349 Int	Intensity : 208,231

Column	Cartridge 2. / μ m, 5 x 2.1 mm					
	(cat.# 9309A0252)					
Mobile	5 mM ammonium formate in water with					
phase A	0,1% formic acid					
Mobile	5 mM ammonium formate in methanol					
phase B	with 0,1% formic acid					
	Time (min.)	Flow	%B			
		(mL/min.)				
Time	2.20	0.45	30			
Program	2.40	0.45	50			
	8.20	0.45	70			
	11.20	0.45	75			
	12.20	0.45	90			
	12.60	0.45	90			
	12.61	0.45	75			
	13.20	0.45	75			
	13.21	0.45	30			
	16	0.45	30			
Oven Ter	np.	40oC				
Sample T	lemp.	15oC				
Inj. Volut	ne	10 μL				

 225000
 1:Alfatoxin G2 TIC(+)
 2:Alfatoxin G1 TIC(+)
 3:Alfatoxin B2 TIC(+)

 1:Alfatoxin G2 331.0000>313.2000(+)
 E: -24.0 2:Alfatoxin G1 329.0000>243.2000(+).4Eat@6r0B2 315.1000>287.2000(+) CE: -25.0

 200000
 1:Alfatoxin G2 331.0000>189.2000(+)
 E: -24.0 2:Alfatoxin G1 329.0000>200.2000(+).4Eat@6r0B2 315.1000>287.2000(+) CE: -25.0

 1:Alfatoxin G2 331.0000>189.2000(+)
 E: -42.0 2:Alfatoxin G1 329.0000>200.2000(+).4Eat@6r0B2 315.1000>259.2000(+) CE: -29.0

 1:Alfatoxin G2 331.0000>245.2000(+)
 E: -42.0 2:Alfatoxin G1 329.0000>200.2000(+).4Eat@6r0B2 315.1000>259.2000(+) CE: -29.0

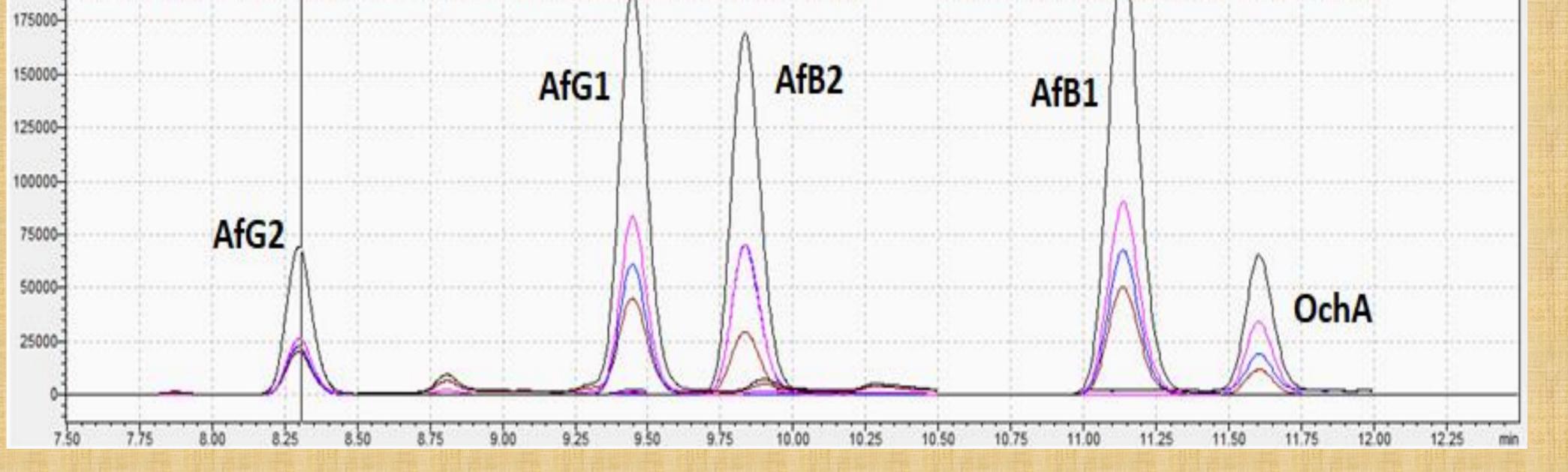
 1:Alfatoxin G2 331.0000>245.2000(+)
 E: -30.0 2:Alfatoxin G1 329.0000>311.2000(+).4Eat@6r0B2 315.1000>243.2000(+) CE: -42.0

MS/MS	Shimadzu LCMS-8045
Ion Mode	ESI+

Table 1. Chromatographic conditions

Analyte	Precursor Ion	Product Ion Quantifier	Product Ion Qualifier
AfG2	331.0	189.2	313.2
AfG1	329.0	200.2	243.2
AfB2	315.1	287.2	243.2
AfB1	312.9	285.2	241.2
OchA	404.1	239.1	358.2

Table 2. Analyte transition



4:Alfatoxin B1 TIC(+) 5:Ochratoxin A TIC(+)

4:Alfatoxin 81 312.90005236(2000(#)-C80-23000>239.1000(+) CE: -22.0

4:Alfatoxin B1 313.0000523602000(+)4C8040000>358.2000(+) CE: -13.0

4:Alfatoxin B1 312.90005286(2800(n) 4:80-84000>221.1000(+) CE: -35.0

Figure 1. Typical chromatograms of aflatoxins

CONCLUSION A novel LC/MS/MS method was developed and validated for determination of aflatoxins and ochratoxin A in cannabis flowers and extracts