

RAMAN SPECTROSCOPY FOR CHARACTERIZATION OF PLANT BIOACTIVE COMPONENTS USED AS NUTRACEUTICALS

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Background: Infrared (IR) and Raman spectroscopy are complementary vibrational spectroscopy techniques, which may provide important composition-related informations of complex plant/food samples. Generally, vibrational measurements can be performed directly on plant tissues or on samples isolated from the plant material by distillation or extraction. Evaluation of biological tissues without extraction, which can lead to degradation of the bioactive components (ex., antioxidants), short time of analysis, a high degree of precision, use in order to perform fast quality checks of raw materials or continuous controlling of the production, are advances of application of Raman spectroscopy to analysis of nutraceutical compounds.

Materials and methods:

This technique allow to obtain spectra (Raman fingerprints) which present characteristic key Raman bands of individual bioactive components (Table 1,2 and 3). These bands provide information about the chemical composition of the investigated samples as primary (proteins and amino acids, lipids and fatty acids, carbohydrates) and secondary metabolites (flavonoids, polyphenols and other phenolic substances, terpenoids (mono-, sesqu-, and tetraterpenes), alkaloids, nitrile compounds, iridoids) present.

Table 1: Selected Characteristic Raman Vibrational Modes **Resulting From Plant Lipids and Carbohydrates***

| Analyte | Wave Number (cm ⁻¹) | Vibrational Mode |
|--------------------|------------------------------------|-----------------------|
| Lipids/Fatty acids | 3008 | =C-H |
| | 2970 | -CH ₃ |
| | 2940 | $=CH_2$ |
| | 1670 | C=C trans |
| | 1660 | C=C cis |
| Carbohydrates | | |
| α-Glucose | 847 | (C–O–C) skeletal mode |
| β-Glucose | 898 | (C–O–C) skeletal mode |
| β-Fructose | 868 | (C–O–C) skeletal mode |
| Sucrose | 1462 | d(CH ₂) |
| Maltose | 847 | (C–O–C) skeletal mode |

Table 2: Selected Characteristic Raman Vibrational Modes Resulting From Plant Proteins*

| Analyte | Wave Number (cm ⁻¹) | Vibrational Mode |
|---------------|------------------------------------|---|
| Cystine | 510 | S-S stretch |
| Cystine | 525 | S-S stretch |
| Methionine | 630–670 700–745 2550–2580 | C-S stretch C-S stretch S-H stretch |
| Tyrosine | 850/830 | Resonance between ring fundamental and overtone |
| Tryptophan | 760, 880, 1360 | Indol ring |
| Phenylalanine | 1006 | Ring breathe |
| Histidine | 1409 | N-Deuteroimidazole |
| Aspartic acid | 1400–1430 | C=O stretch of carboxyl group |
| Glutamic acid | 1700–1750 | C=O stretch of carboxyl or ester group |
| Amide I | 1655–1685 | Amide C=O stretch, N-H wagging |
| Amide III | 1235–1280 | N-H in-plane bend, C-N stretch |

CONCLUSION

The ability for rapid monitoring of various plant bioactive components makes Raman spectroscopy one of techniques with future more wide application in the nutraceutical field. As the existed demand to solve complex issues of nutraceuticals is increased recently, investigation of the changes in the functionality of these specific substances with the addition or loss of nutraceutical compounds (both in foods and in model) Systems) by using multidisciplinary approach, Raman Table 3: Selected Characteristic Raman Vibrational Modes **Resulting From Some Plant Terpene Compounds***

| Wave Number (cm ⁻¹) | Vibrational Mode |
|------------------------------------|---|
| 1674; 1382 | C=C; -CH ₃ |
| 1679 | C=C |
| 1678 1645 | (cyclohexane C=C) (ethylene C=C) |
| 1659 666 | C=C δ(ring) |
| 1677 1436 | C=C δ(CH ₂) |
| 1671; 1632 | C=C |
| 1524; 1157 | C=C; C-C |
| 1527; 1157 | C=C; C-C |
| 1510; 1156 | C=C; C-C |
| | Wave Number (cm ⁻¹)1674; 138216791679164516451659666167714361524; 11571527; 11571510; 1156 |

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