



ISSN 2348 - 0319

Journal home page: <http://www.journalijar.com>

INTERNATIONAL JOURNAL  
OF INNOVATIVE AND  
APPLIED RESEARCH

## RESEARCH ARTICLE

## IMAGING OF DEEP VENOUS THROMBOSIS USING RADIOACTIVE LABELED TIROFIBAN: ANIMAL MODEL EVALUATION

Marija Darkovska Serafimovska<sup>1</sup>, Emilija Janevik-Ivanovska<sup>1</sup>, Icko Djorgoski,<sup>2</sup> Nenad Ugresic<sup>3</sup>

1. Goce Delcev University, Faculty of Medical Sciences, Stip, Republic of Macedonia.

2. University St. Cyril and Methodius, Faculty of Natural Science and Mathematics, Skopje, Republic of Macedonia.

3. University of Belgrade, Faculty of Pharmacy, Belgrade, Serbia.

**Abstract:**

Imaging of acute thrombus, especially the very prevalent condition of acute deep vein thrombosis is usually relied on conventional imaging techniques utilizing either ultrasonography or contrast venography. The former procedure is limited by accuracy and the latter by technical considerations. Recent advances in the understanding of the pathogenesis of acute clot at the molecular level have suggested new opportunities for detection of the acute thrombotic process based on the biomolecular behavior of components of the clotting process including the formed element of the blood, the platelet.

Thus, development of radiolabelled small peptide or peptidomimetic ligands that can bind platelets and their specific expressed receptor have been suggested as a new approach to detect clot location and, more essentially, determine the age and morphology of the evolving thrombus. This new approach has focused on the use of a series of radiolabelled platelet GPIIb/IIIa receptor antagonists.

Tirofiban N-(butylsulfonyl)-4-O-(4-(4-piperidyl)-L-tyrosine is a non-peptide tyrosine derivative. The aim of the study was to introduce radioactive labelled tirofiban as a specific imaging agent for acute DVT.

The labeling was performed with Technetium-99 in the presence of a stannous reducing agent. The labelled preparation showed a fast blood clearance in the normal rat model (without induced thrombosis). More than 80% of the injected dose was eliminated from the circulation in the first hour after injection. Biodistribution and visualization of the labelled molecule was carried out using an experimental model of thrombosis in the male Wistar rat. Planar images were obtained 30 min and 60 min after application of  $2 \times 10^6$  imp/min <sup>99m</sup>Tc-tirofiban, in the rat's tail vein. Sensitivity and specificity were determined using the ratio 'left leg positive for DVT' and 'right leg negative for DVT'. The obtained ratio was 1.54 after 30 min and 5.04 after 60 min. These values were considered as positive in the detection of acute DVT. The high DVT uptake shows that radiolabelled tirofiban in the introduced rat model can be the promising agent for imaging of deep venous thrombosis.

**Key Words:** Radionuclide labeling, Imaging, Deep Venous Thrombosis; <sup>99m</sup>Tc-Tirofiban, Rat model

**Introduction**

Venous thromboembolism is a complex vascular disease with a multifactorial pathogenesis that results in significant morbidity and mortality. The first and more common manifestation is deep venous thrombosis (DVT), which usually arises in the deep veins of the calf and spreads upwards. Pulmonary embolism, the second and more serious manifestation, occurs as a complication of DVT proximal to the deep calf veins [1].

Because the incidence of deep venous thrombosis (DVT) and pulmonary embolism (PE), in high-risk populations, in hospitalized patients, as well as in the general population remains is still remarkably high, yet an accurate diagnosis of DVT and PE continues to be unreliable [2,3].

All available imaging procedures (duplex ultrasound, magnetic resonance and contrast venography as a standard test for validating new diagnostic procedures) do not reflect the metabolic activity of the clot and therefore they may overestimate the presence of active clots [4]. Only nuclear medical examinations can provide an image that includes information on thrombus formation [25]. The development of radioactive agents for scintigraphic imaging of DVT and PE is centered on the use of <sup>99m</sup>Tc-labeled peptides or peptidomimetics specific for resting or activated platelets [4-9]. They are smaller in size, easier to produce than monoclonal antibodies, expected to clear more rapidly from circulation than radiolabeled proteins, are less likely to induce any immunologic reaction, and yet, in most cases, they enjoy receptor specificity and binding constants as high as those of the monoclonal antibodies.

<sup>99m</sup>Tc is a radionuclide inexpensive, easy to obtain worldwide, and decays with emission of  $\gamma$ -rays (140 keV, 90%) that can be efficiently detected by  $\gamma$ -cameras. Its half-life (6 h) is long enough to perform examinations before excessive radioactive decay has occurred, yet not too long to persist in the body long after the examinations have been performed.

Our approach was focused on the use of a series of radiolabelled platelet GPIIb/IIIa receptor antagonists [6, 10]. The GPIIb/IIIa receptors are expressed on the membrane surface of activated platelets and play an integral role in platelet aggregation and thrombus formation [10]. Initial actions in thrombus formation frequently involve the activation of platelets by thrombogenic conditions and their subsequent aggregation. Platelet aggregation is mediated by fibrinogen, which binds via the Arg-Gly-Asp (RGD) sequence to the GPIIb/IIIa receptor expressed on activated platelets. Since the GPIIb/IIIa receptors are expressed only on the membrane surface of activated platelets, with 50 000–90 000 GPIIb/IIIa binding sites per platelet [2], the GPIIb/IIIa receptor makes an excellent target for the development of an imaging agent than bound with high specificity to activated rather than to unactivated platelets. They would be differentially incorporated in the thrombus (activated platelets) and the circulating platelets (resting or relatively less activated) [11, 12].

These molecules represent glycoprotein (GPIIb/IIIa) receptor antagonists and they act as true biochemical markers of active thrombosis [10, 11].

One peptide, <sup>99m</sup>Tc-P280 (5), is approved by the Food and Drug Administration under the tradename AcuTect (<sup>99m</sup>Tc-P280; Diatide, Inc.) (8), which can image acute thrombi but not old clots or PE.

The goal of our work was to use the small non-peptide derivative or peptidomimetic ligands with high specificity for the GPIIb/IIIa receptor and incorporate a convenient radionuclide for imaging purposes was initiated from the already existing data presented using labelled peptides and GPIIb/IIIa receptor antagonists in the diagnosis of acute DVT. One promising GPIIb/IIIa receptor antagonist is tirofiban (Aggrastat<sup>TM</sup>, Merck, Inc.), non-peptide tyrosine derivative that inhibits fibrinogen binding [14]. Tirofiban hydrochloride monohydrate is chemically described as N-(butylsulfonyl)-O-(4-[4-piperidinyl]butyl)-L-tyrosine monohydrochloride monohydrate (Fig. 1.) The empirical formula of Tirofiban is  $C_{22}H_{36}N_2O_5S \cdot HCl \cdot H_2O$  and molecular weight of 495.08 [2,3].

It is useful in combination with heparin and aspirin in the management of patients with unstable angina or non-Q-wave myocardial infarction, including patients who may subsequently undergo percutaneous transluminal coronary angioplasty (PTCA) [13, 14].

Adjunctive therapy with a GP IIb/IIIa-receptor inhibitor can reduce the incidence of cardiac ischemic events, including subsequent myocardial infarction (MI) and death, in patients with non-ST-segment-elevation acute coronary syndromes [5,6].

When Tirofiban is prescribed and administered to the patient according to the recommended treatment, >90% inhibition is attained by the end of the 30-minute infusion. Platelet aggregation inhibition is reversible following cessation of the infusion of Tirofiban. Tirofiban with a half-life of approximately 2 hours [15] is not strongly bound to plasma protein, and protein binding is concentration-independent in the range of 0.01–25  $\mu\text{g mL}^{-1}$ . The unbound fraction in human plasma is 35%. The distribution volume of Tirofiban in the steady state is about 30 liters.

To obtain the concentration of tirofiban appropriate for labeling as potential imaging radiopharmaceuticals and in the same time not receive therapeutic effect, we have labeled tirofiban with <sup>99m</sup>Tc and evaluated it in vitro and in rats, with experimental venous thrombosis established and modified afterwards, according to the needs of a particular line of research [16, 17].

## Material and Methods

### Materials

#### Tirofiban

The structure of tirofiban (tirofiban hydrochloride monohydrate) N-(butylsulfonyl)-O-[4-(4-piperidinyl)butyl]-L-tyrosine monohydrochloride monohydrate is shown in Fig. 1. Tirofiban hydrochloride monohydrate is a white to off-white, non-hygroscopic, free flowing powder, with a molecular weight of 495.08. It is very slightly soluble in water [2,3].

Isotope- <sup>99m</sup>Tc

<sup>99m</sup>Tc-pertechnetate was obtained from a commercial <sup>99</sup>Mo/<sup>99m</sup>Tc generator (10 GBq) (Schering /CIS-biointernational).

### **Radiolabelling**

Labelling of tirofiban with <sup>99m</sup>Tc

Tirofiban was labelled using the method of direct labelling under nitrogen. The kit freeze dried formulation contains a mixture of tirofiban (20 nmol) dissolved in buffer (ethanol 96%/PBS 0.01M, pH 7.3, 1:3 vol./vol.) and stannous chloride (10 nmol) as a reducing agent. Sodium (<sup>99m</sup>Tc) pertechnetate (specific activity 740–4500 MBq/mL) containing 100 MBq/mL was added and the reaction mixture incubated for 15 min at room temperature.

The quality control was done by paper chromatography and instant thin layer chromatography (ITLC) using two solvents — 95% acetone and saline. The percentage labelled yield was recorded by gamma scanner (Veenstra Instrumenten B.V. VCS-103 V1.06).

### **Blood clearance**

<sup>99m</sup>Tc-tirofiban was injected intravenously into rats. Blood samples were drawn from the previously prepared carotid vein using a sterile syringe at 5, 15, 30, 45 min and 1, 2, 4, 6 and 24 h after injection. All samples were of the same volume and their radioactivity measured in a gamma counter and compared with a standard.

*In vitro* platelet binding

The binding study was performed with rat and human platelets isolated and treated according to protocol for platelet labelling (TROMBO-SCINT). Platelets were incubated for 30 min with tirofiban radiolabelled with <sup>99m</sup>Tc. The percentage of binding was measured after one and two washing steps.

### **Animal studies**

Male Wistar rats weighing 220–250 g were used throughout the study, anaesthetized by intraperitoneal injection of water solution of Nesdonal (concentration 20 mg/kg body weight).

Experimental animal model [18, 19]

Venous thrombosis was induced by ligation of the femoral vein in rats whose blood was made hypercoagulable by intravenous administration of tissue thrombin [17]. For the thrombosis model, the body temperature of the rats was maintained at 37°C. In brief, a short incision was made in the skin and subcutaneous tissue in the left groin region and the femoral neurovascular sheath was gently exposed [20]. An approximately 10 mm long portion of the left femoral vein, distal to the inguinal ligament was isolated by rubbing it against the blade of a pair of forceps and this segment was collapsed. The collapsed segment of femoral vein between the clamps was traumatized by striking and the twenty units of thrombin (in 0.2 mL saline) were injected into the segment with a needle. A semiconstricting ligature was placed downstream to prevent the clot from slipping away.

Imaging

The thrombi developed in the rats were visualized using tirofiban radiolabelled with <sup>99m</sup>Tc. Planar images were obtained 30 min and 60 min after application of  $2 \times 10^6$  counts/min in 50–100 µL <sup>99m</sup>Tc-tirofiban in the rat's tail vein. Syringes should be measured before and after injection in order to determine accurately the radioactivity of material injected. The sensitivity and specificity of the radiopharmaceuticals were determined using the ratio 'left leg positive for DVT' and 'right leg negative for DVT' using the ROI technique.

### **Biodistribution**

The biodistribution studies were carried out using the same experimental model of thrombosis in male Wistar rat and by injection of the radiolabelled products. After the desired time period had elapsed, the animals were sacrificed and the samples of organ of interest (heart, lung, liver, spleen, kidney, thrombotic and normal tissue) were collected and placed in pre-weighed counting tubes. These were then counted in a gamma counter together with a standard prepared from a known dilution of the injected material (preferably prepared at the time of injection). The total activity injected into each rat was determined and the activity remaining in the tail subtracted. The uptake of the labelled products in each tissue was calculated and the specific uptake of the products quantified using ratio 'left leg positive for DVT' and 'right leg negative for DVT'.

## **Results**

This study used a rat model of DVT to evaluate radiolabelled tirofiban, platelets GPIIb/IIIa antagonist for its potential use in the detection of rapidly growing venous thrombi. Two different methods of labelling were selected using <sup>99m</sup>Tc as radioisotope.

Labelling of tirofiban with <sup>99m</sup>Tc

The specific activity of the radiolabelled product was  $9.2 \times 10^{18}$  to  $1.0 \times 10^{19}$  counts/min/mol.

The percentage of obtained complex after labelling was more than 95% (in acetone), more than 85% (in saline), with the free pertechnetate (<sup>99m</sup>TcO<sub>4</sub><sup>-</sup>) less than 5% (Fig. 2).

The labelled product was stable without changing the percentage of labelling after 2 h at room temperature.

### In vitro platelet binding

### Blood clearance

## Imaging studies

### Biodistribution studies

Biological distribution data for the labelled 99mTechnetium-tirofiban at different times after IV administration are presented in Fig. 6. These biodistribution studies demonstrated some different behavior. Results obtained from the radioactivity distribution of 99mTechnetium-tirofiban at 30 min and 60 min after IV administration clearly present a the accumulation in the critical organs. This is in accordance with the presence of destroyed platelets carrying radioactivetirofiban. However, the radiolabelled product was rapidly cleared from the circulation as represented by a low percentage of radioactivity in the heart and lung. The obtained ratio 'left leg positive for DVT' and 'right leg negative for DVT' was 1.54 after 30 min and 5.04 after 60 min for the tirofiban labelled with technetium (Fig. 7). These values were considered as positive in the detection of acute DVT.

## FIGURES

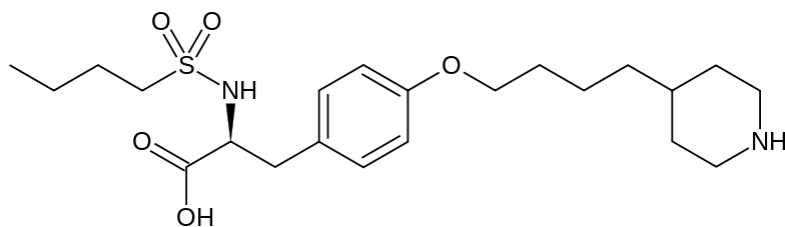
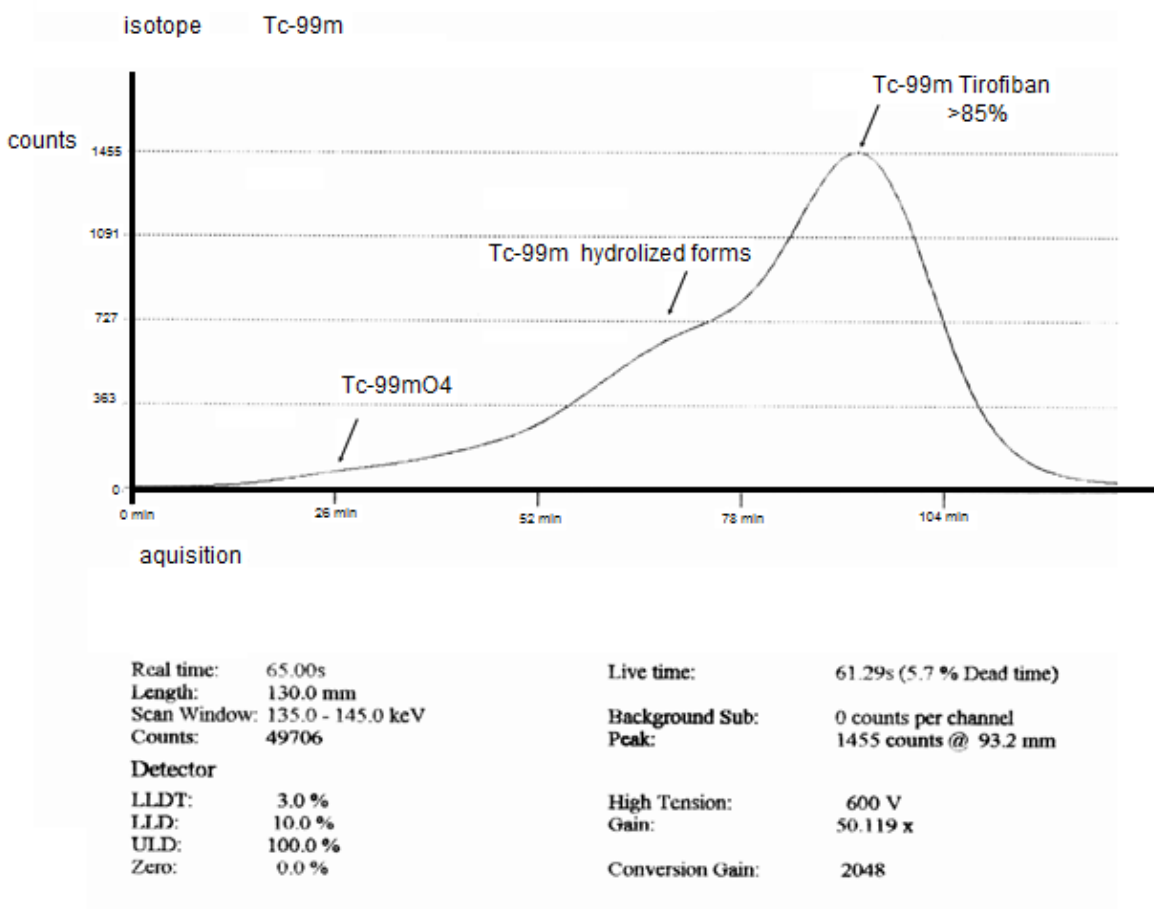


Figure 1. Chemical structure of Tirofiban hydrochloride.

a.



b.

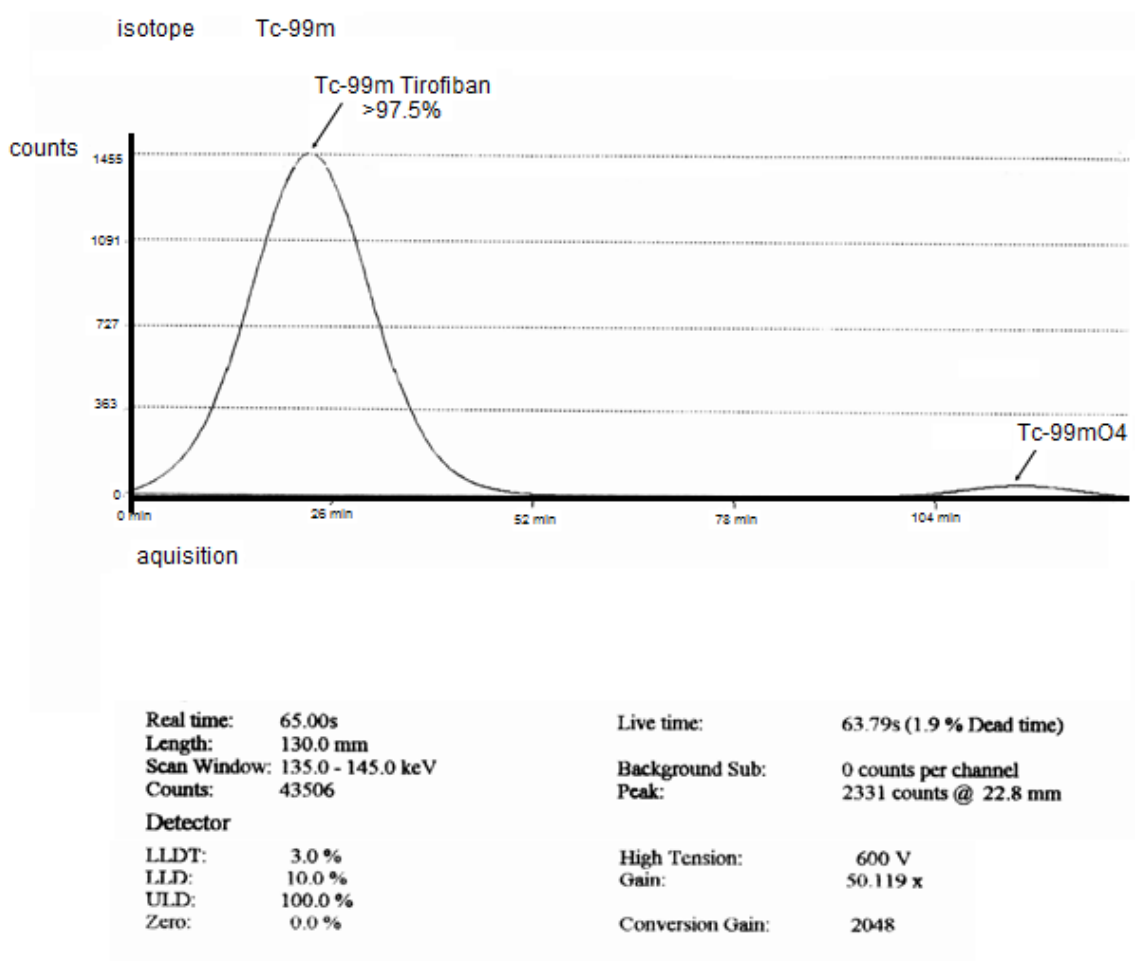


Figure 2.ITLC radiochromatograms of 99mTc-tirofiban in (a) 95% acetone (b) saline.

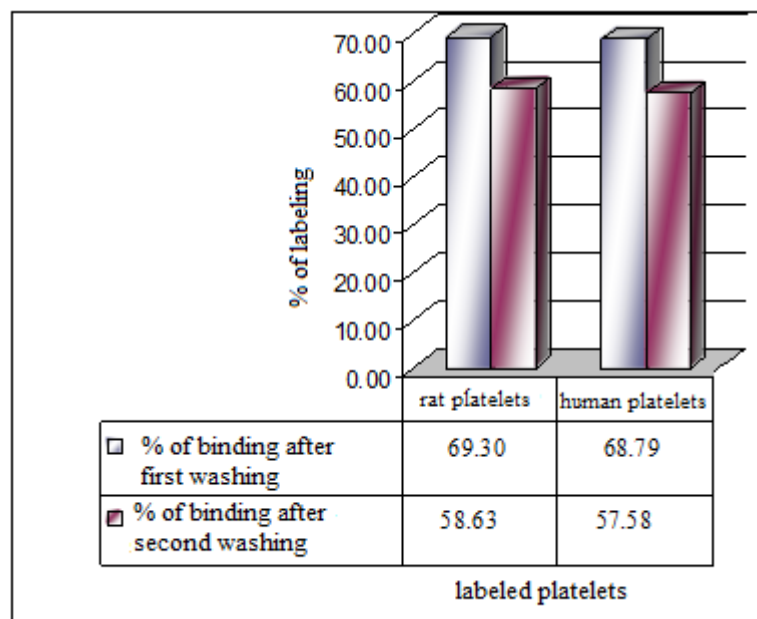


Figure 3. In vitro binding of rat and human platelets isolated and treated with radiolabelled tirofiban.

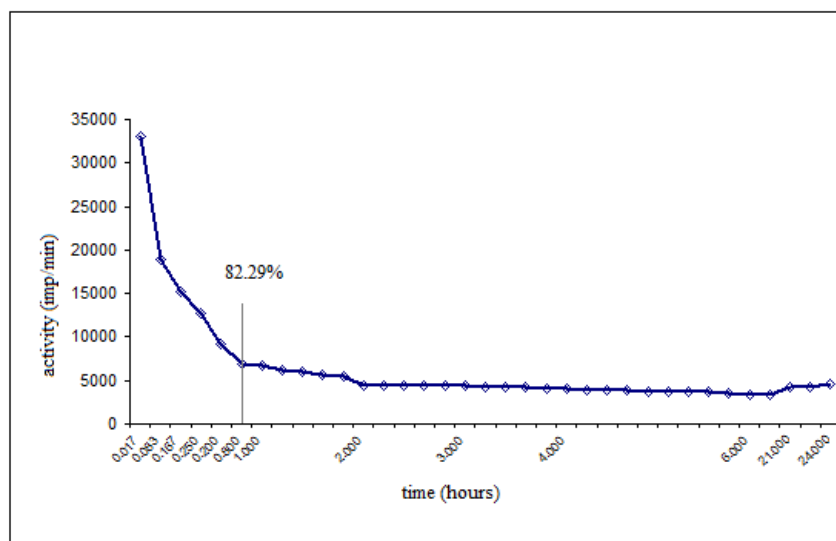


Figure 4. Blood clearance of radiolabelled tirofiban -  $^{99m}\text{Tc}$ -tirofiban in normal rat



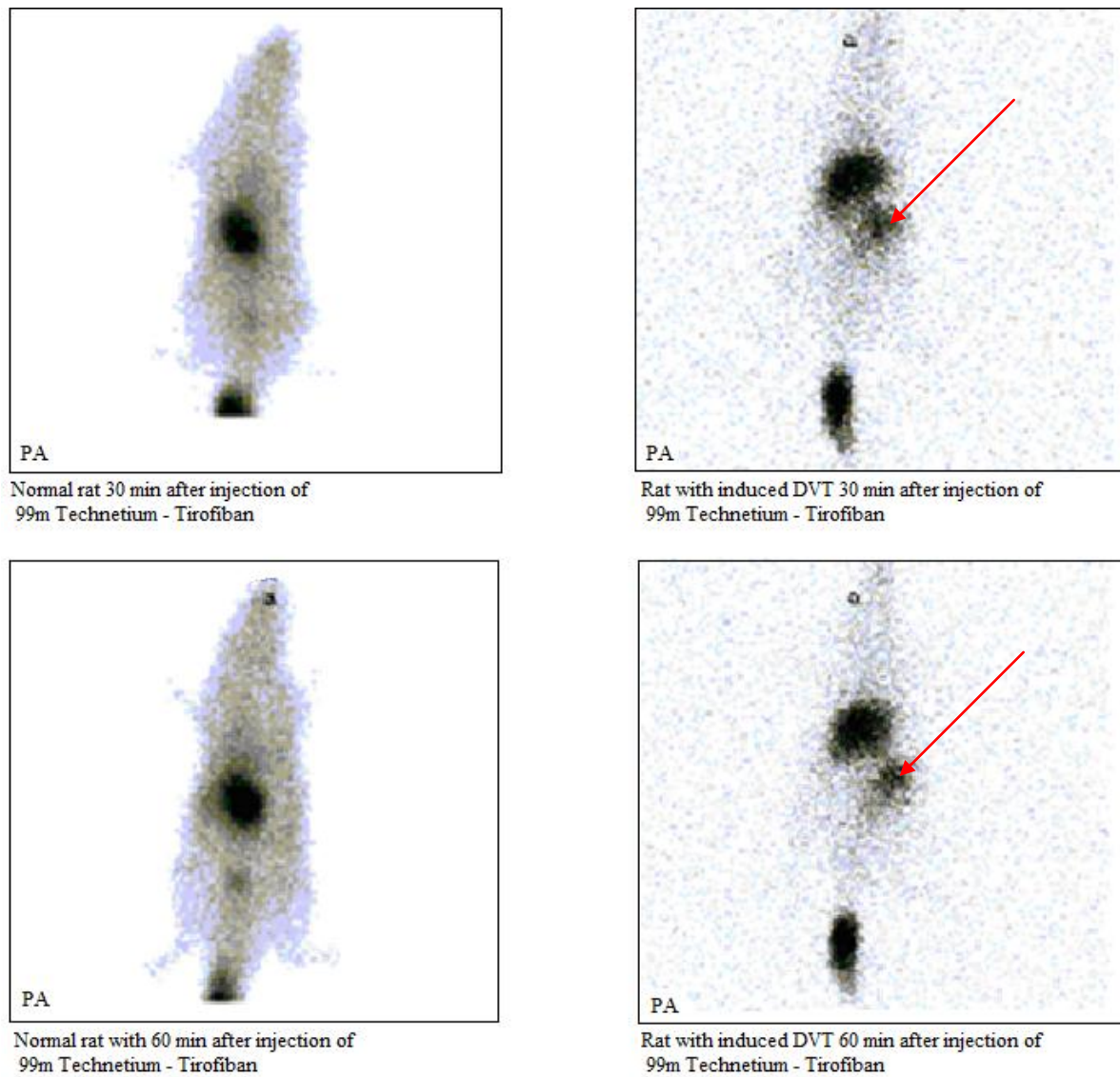


Figure 5. Gamma camera images after IV injection of  $^{99m}\text{Tc}$ -tirofiban in rats



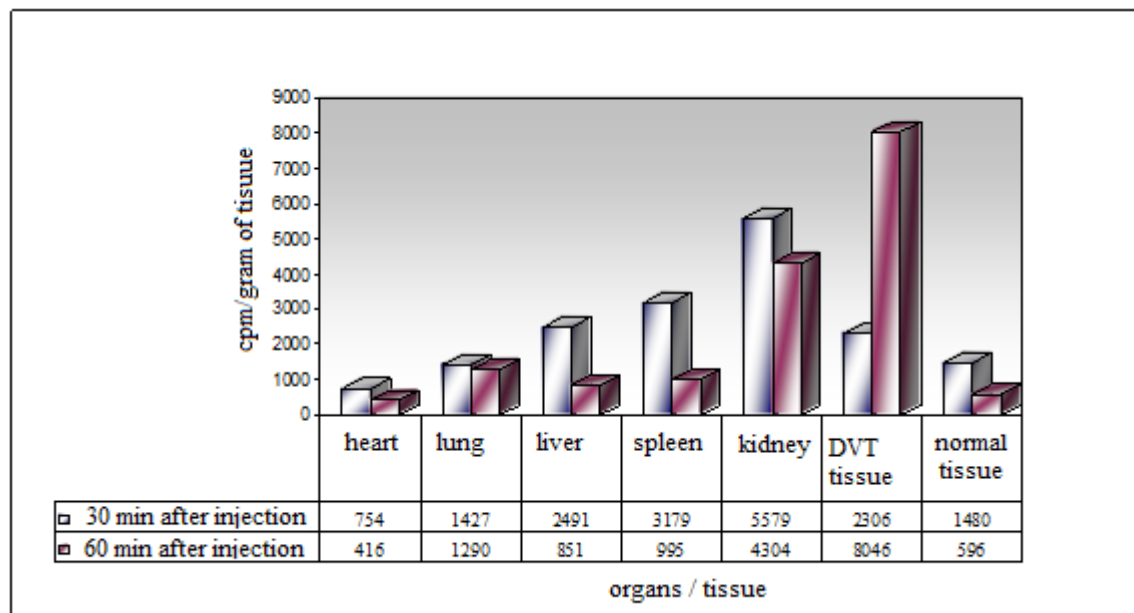


Figure 6. Biodistribution studies in rat with induced DVT and after injection of <sup>99m</sup>Technetium labeled tirofiban

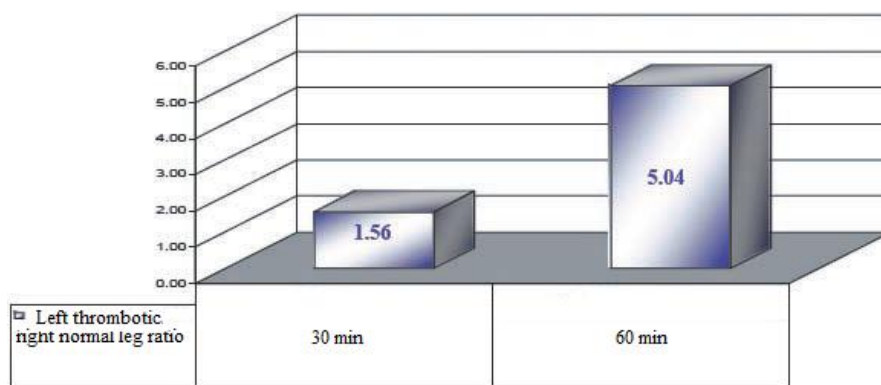


Figure 7. Left thrombotic/right normal leg ratio using ROI technique from gamma camera images after injection of <sup>99m</sup>Technetium-tirofiban.

## DISCUSSION

Blood coagulation has long been considered an important factor in the pathogenesis of venous thrombosis. The relative contribution of stasis, altered coagulability of the blood, vessel wall damage and circulation of leucocytes or platelets to the pathogenesis of venous thrombosis remains in dispute. On the other hand, it is known that a hypercoagulable state contributes significantly to the thrombotic process. Animal models based on these activation mechanisms have been designed as a means of studying potential imaging agents for diagnosis of acute DVT. The venous stasis thrombosis model consists of inducing blood stasis in the femoral vein of rats after prior injection of a procoagulant. A radionuclide imaging agent that binds to platelets being incorporated into an active thrombus but one which, if not bound, clears rapidly from the blood would have great potential for acute DVT detection.

Platelets expressed from the cell surface of GPIIb/IIIa receptors undergo the conformational change that makes them available for binding fibrinogen. Cross-linkage of activated platelets by the bivalent fibrinogen molecule to form a hemostatic plug is the primary episode of thrombosis. An imaging agent, ideally labelled with <sup>99m</sup>Tc and capable of binding actively and specifically to the GPIIb/IIIa receptor on activated platelets, would give images of active or acute venous thrombosis.

The aim of the study was to evaluate tirofiban as a specific imaging agent to GPIIb/IIIa receptors in the case of experimentally induced acute DVT in the rat experimental model. Radionuclide imaging offers considerable potential as a successful diagnostic agent which would address a few important criteria, i.e. rapid, non-invasive, cost effective and accurate. The published data [7, 13, 19] indicate that radiolabelled tirofiban binds preferentially to GPIIb/IIIa receptors on activated platelets and can differentiate acute DVT from chronic venous thrombosis. Radiolabelled tirofiban is a functional rather than an anatomical imaging modality. The use of radiotracers allows an understanding of the bioavailability process, biodistribution and kinetics of any molecule labelled with an isotope, a procedure which does not alter the molecule's biological properties. In the current work, technetium (to evaluate tirofiban as a specific imaging agent to GPIIb/IIIa receptor in the case of experimental induced acute DVT in the rat experimental model) was chosen as a radiotracer for biodistribution and imaging studies in the experimental thrombosis induced in rats for its favourable radiation and physical characteristics, ready availability, possibility of labeling and low cost.

Commercially available technetium-apcitide (AcuTect), previously known as 99mTc-P280, which binds to the GPIIb/IIIa receptor, is the first imaging agent used in the clinical studies to detect acute DVT [21, 22]. In this study, although the 99mTc-apcitide images obtained at 2 h after tracer injections show the greatest overall accuracy in comparison with earlier images, combined analysis of image sets from at least two time points (30 min and 60 min) provides greater accuracy in the detection of acute DVT in the patients. These results correlate with data obtained from the animal model. 99mTc-technetium-tirofiban is accurate in the detection of acute DVT, especially 60 min after application. The results obtained from the animal design experimental studies showed that the ratios 'left leg positive for DVT' and 'right leg negative for DVT' for the radiolabelled preparation of tirofiban are well within the range of that expected for a successful imaging agent.

## Conclusion

Animal models provide convenient screening tools for radiolabelled products before a new radiopharmaceutical is further developed in clinical trials [23].

The tirofiban labelled with 99mTc-technetium was found to have good thrombus uptake in vivo experiments. Obtained data indicate that radiolabelled tirofiban binds preferentially to GPIIb/IIIa receptors on activated platelets and can differentiate acute DVT from chronic venous thrombosis.

These results can be helpful in the further clinical investigation of patients with acute DVT.

## References

1. Hyers TM, Venous thromboembolism, Am. J. Respir. Crit. Care Med. 1999; 159: 1–14.
2. Horlander KT, Mannino DM, Leeper KV. Pulmonary mortality in the United States, 1979–1998. Arch Intern Med 2003; 163:1711–1721.
3. Medicure Pharma. Aggrastat (Tirofiban hydrochloride) injection premixed and injection prescribing information. Somerset, NJ; 2007 Nov.
4. Anon. Tirofiban hydrochloride. Drugs Future. 1995; 20:897-901.
5. Aruva MR, Daviau J, Sharma SS, Thakur ML. Imaging Thromboembolism with Fibrin-Avid 99mTc-Peptide: Evaluation in Swine. J Nucl Med. 2006; 47(1): 155–162.
6. Knight LC, Radcliffe R, Maurer AH, et al. Thrombus imaging with 99mTc synthetic peptides based upon the binding domain of a monoclonal antibody to activated platelets. J Nucl Med 1994; 35:282–288.
7. Knight LC, Maurer AH, Romano JE. Comparison of iodine-123-disintegrins for imaging thrombi and emboli in a canine model. J Nucl Med 1996; 37:476–482.
8. Pearson DA, Lister-James J, McBride WJ, et al. Thrombus imaging using 99mTc labeled high potency GPIIb/IIIa receptor antagonist: chemistry and initial biological studies. J Med Chem 1990; 39:1372–1382.
9. Lister-James J, Vallabhajosula S, Moyer BR, et al. Pre-clinical evaluation of technetium-99m platelet receptor-binding peptide. J Nucl Med 1997; 38:105–111.
10. Line BR, Cran P, Lazewatsky J, et al. Phase I trial of DMP444, a new thrombus imaging agent [abstract]. J Nucl Med 1996; 37(suppl):117P.
11. Barrett JA, Damphousse DJ, Heminway SJ, et al. Biological evaluation of 99mTc cyclic glycoprotein IIb/IIIa receptor antagonists in the canine arteriovenous shunt and deep vein thrombosis models: effects of chelators on biological properties of [99mTc] chelator–peptide conjugates. Bioconjug Chem 1996; 7:203–208.

12. Collier BS, Cell adhesion in vascular biology: Platelet GPIIb/IIIa antagonists: The first anti-integrin receptor therapeutics, *J. Clin. Invest.* 1997; 99: 1467–1471.
13. Verstraete M. Synthetic inhibitors of platelet glycoprotein IIb/IIIa in clinical development, *Circulation* . 2000; 102: e76–e80.
14. Weiss DJ. et al., Platelet kinetics in dogs treated with a glycoprotein IIb/IIIa peptide antagonist, *Toxicol. Pathol.* 2000; 28: 310–316.
15. ICH Topic Q 2 (R1) Note for guidance on validation of analytical procedures: Text and methodology (CPMP/ICH/381/95), European Medicines Agency, London, 1995
16. Ranjitha KS, Rao AL. Development and validation of new RP-HPLC method for determination of Tirofiban in pharmaceutical formulation. *IJPCBC*. 2011; 1 (1): 43-47
17. [http://www.accessdata.fda.gov/drugsatfda\\_docs/label/2012/020912s018,020913s017lbl.pdf](http://www.accessdata.fda.gov/drugsatfda_docs/label/2012/020912s018,020913s017lbl.pdf) assessed on 14<sup>th</sup> of July, 2012.
18. Callas JW, FAREED DDJ. A survey of animal models to develop new and novel antithrombotic agents, *New Therapeutic Agents in Thrombosis and Thrombolysis* (SASAHARA, A.A., LOSCALZO, J.L., Eds) 1997; 9–28.
19. Herbert JM, Bernat A, Maffrand JP. Importance of platelets in experimental venous thrombosis in the rat, *Blood*. 1992; 80(9): 2281–2286.
20. Lister-Jones J, Mauer A. Thrombus imaging with a technetium 99m labeled, activated platelet receptor binding peptide. *J. Nucl. Med.* 1996; 213: 207.
21. Olfert ED, Cross BM, McWilliam AA. (Eds), *Guide to the care and use of experimental animals*, Canadian Council on Animal Care. 1993; Vol. 1, 2nd ed.
22. Stassen JM. Animal model to assess the safety and efficacy of thrombolytic agents. *New Therapeutic Agents in Thrombosis and Thrombolysis* (Sasahara AA, Loscalzo J.L. Eds), Streiff MB, Vena 1997; 451–474.
23. Zhou Y, Chakraborty S, Liu S. Radiolabeled Cyclic RGD Peptides as Radiotracers for Imaging Tumors and Thrombosis by SPECT. *Theranostics* 2011; 1: 58-82
24. Carretta RF, Streek PV, Weiland FL. Optimizing Images of Acute Deep-Vein Thrombosis Using Technetium-99m-Apcitide. *J Nucl Med Techn.* 1999; 27: 271-275.
25. Houshmand S, Salavati A, Hess S, Ravina M, Alavi A. The role of molecular imaging in diagnosis of deep vein thrombosis. *Am J Nucl Med Mol Imaging* 2014; 4(5): 406-425